# Atherosclerosis-on-a-Chip

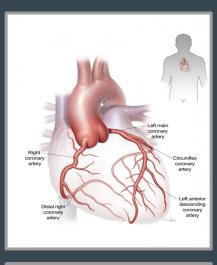
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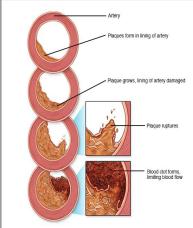
Khadar, Shivansh, Viraj, Vivek, Jackie

# **Introduction & Background**

### **Disease Background**

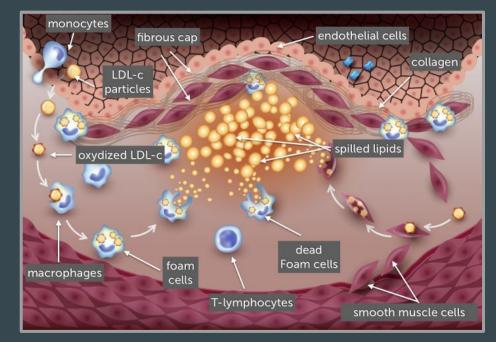
- **Atherosclerosis** Plaque buildup in the arteries that perfuse the heart
  - Diabetes, diet, and lack of exercise are major contributors
- Complications
  - Arrhythmias
  - Heart Failure
  - Heart Attack / Stroke
- Disease Prevalence
  - 18.2 million adults > 20 years old have CAD
  - 360,900 deaths in 2019 from CAD
- Symptoms
  - Angina
  - Shortness of Breath
  - Arm/Shoulder Pain





### **Atherosclerosis: Formation of Subendothelial Fatty Plaques**

- 1. **Vascular insult** occurs to the tunica intima
- 2. **Monocytes migrate** to damaged subendothelial space
- 3. Monocytes take up oxidized LDL becoming foam cells
- 4. **T cells become activated** releasing cytokines
- 5. **Growth factors** activate smooth muscle cells
- 6. **Smooth muscle cells** take up LDL and deposit at the injury site resulting in subendothelial plaque



### Target Physiology for OOC: Coronary Arteries

#### • Three Key Layers

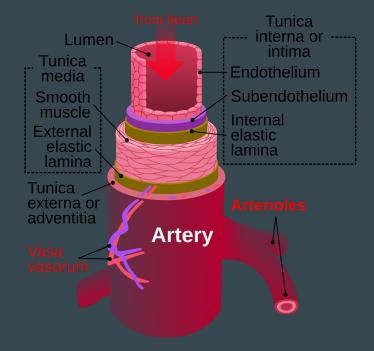
- Tunica Intima
- Tunica Media
- Tunica Externa/Adventitia

#### • Multiple Different Cell Types

- Epithelial Cells
- Smooth Muscle Cells
- Perivascular Adipose Tissue Cells
- Fibroblasts

#### • Physiological Flow Through Lumen

- Flow of Cells Across Tunica Intima for Disease Pathophysiology
- Expansion of Tunica Intima and Narrowing of Vessel Lumen under Disease State



### Organ-on-a-Chip (OOC)



- Multi-channel 3-D microfluidic cell culture
- Microdevices engineered to contain (human) cells and tissues and to model or mimic organ structures for *in vitro* study
- Highly used in drug discovery and development process to bring the gap between cell study and clinical study closer
- Three main characteristics:
  - 3D nature and arrangements of the tissues on the platforms
  - Presence and integration of multiple cell types to reflect a more physiological balance of cells
  - Presence of biomechanical forces relevant to the tissue being modelled

### **Advantages and Limitations of OOC Technology**

#### Pros:



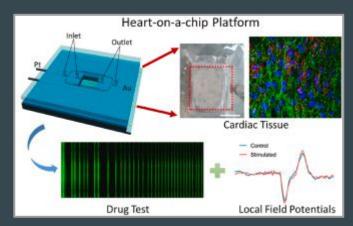
- Cellular and specific tissue architecture is well maintained to emulate chemical gradients and biomechanical forces
- Can vascularized or perfuse tissues to bring nutrients and fluidic flow to cells
- Can incorporate real-time tissue function sensors

#### Cons:

- Air bubbles can form easily in the channels of devices and interrupt the physiological response of the cells
- Extracellular matrix tend to degrade as time goes on
- PDMS can absorb organic compounds and cause a nutritional imbalance
- Organ volume versus organ area imbalance effect on mimicking function of original organ is controversial

### **Current Technology**

- Comparable Heart-on-a-Chip (HOC) Technology
  - Goal: Reproduce mechanisms of the heart; mimicking mechanics of the heart as a whole
  - Technology exists; more work needed to reproduce highly controlled microfluidic environment
  - Limitations:
    - Material Selection
    - Interface Development
    - Improper Vascularization and Perfusion
- Only a few **specific applications** to **localized heart mechanics**



Many current applications attempt to mimic contractility and electrophysiology of the heart

### **Purpose of Our Atherosclerosis-on-a-Chip**

#### 1. Improvement on Current Fabrication Techniques

- a. Lower Cost
- b. Increased Production Efficiency
- c. Increased Scalability > Increased Accessibility

#### 2. Drug and Toxicology Studies for CAD

- a. Test New Drugs Meant for Other Diseases
  - i. Ensure No Effect on Patient's Pre-Existing CAD
- b. Test Newly Developed Drugs for CAD

#### 3. Highly Patient-Specific Model of the Coronary Arteries

- a. Investigate/Quantify Effects of Other Risk Factors
  - i. Smoking
  - ii. Increased Blood Sugar from Diabetes
  - iii. Diet

### Design of Atherosclerosis-on-a-Chip (AOC)



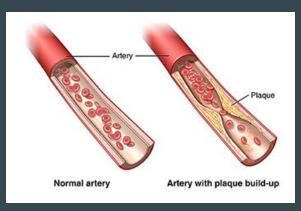
### **Overall Chip Design**

#### • Bioprinting

- Techniques to combine cells and various
   biomaterials to fabricate biomedical parts
  - Aim of imitating natural tissue characteristics
- In our application, we will use bioprinting to create the structure of the coronary arteries for incorporation into OOC
- Microfluidic Chip Development
  - Module-Based
    - Including active and passive mixing modules
  - Fabricated from PDMS

Inducing Atherosclerosis

- Low-density lipoproteins into the endothelial cell layer
- Increasing heart rate with pulsatile flow
- Observe plaque buildup within vessel walls



# **3D Bioprinting**

### **Design Parameters for 3D Bioprinting**

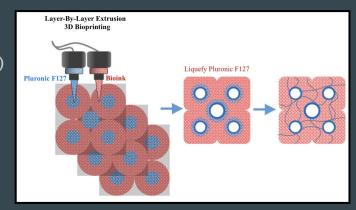
	3D Bioprinting Method	3D Bioprinting Bioink	3D Bioprinting Cells/Materials	
Component(s)	<ul> <li>Pneumatic Microextrusion 3D Bioprinter</li> <li>Coaxial Nozzles for Simultaneous Printing and Crosslinking Material</li> <li>UV Light Source for Photocrosslinking</li> </ul>	<ul> <li>Pluronic<sup>®</sup> F127 (Sacrificial Material)</li> <li>Methacrylated Gelatin (GelMA), Alginate, PEGTA Blend (Cell-Seeded Layer)</li> <li>Methacrylated Gelatin (GelMA), Collagen I, PEGTA Blend</li> </ul>	<ul> <li>Human Umbilical Cord-Derived Vein Cells</li> <li>Human Umbilical-Cord Derived Arterial Cells</li> <li>Mesenchymal Stem Cells</li> <li>Vascular Smooth Muscle Cells</li> <li>NIH 3T3 Fibroblast Cells</li> </ul>	Organ or tissue of interest       3D imaging (optical, MRI, CT)       3D CAD model       Generation of G-co         Organ or tissue of interest
Reasoning	Layer-by-layer microextrusion bioprinting methods, components will be simultaneously printed and crosslinked through multiple nozzles	Pluronic <sup>®</sup> F127 as a sacrificial material for inner lumen; reverse gelation properties Bioink blends for cell growth and proliferation	Different cell types embedded after co-culture with blends to facilitate development of target physiology	

calls

### **Bioprinting Methods**

# • Three Key Layers with Separate Bioinks and Nozzles

- Tunica Intima (Endothelial Layer)
  - GelMA/Hyaluronic Acid/PEGTA (UV Light & CaCl<sub>2</sub>)
  - HUAEC/HUVEC Cells
  - Mesenchymal Stem Cells
- Tunica Media (Smooth Muscle Layer)
  - GelMA/Alginate/PEGTA (UV Light & CaCl<sub>2</sub>)
  - Vascular Smooth Muscle Cells
  - Mesenchymal Stem Cells
- Tunica Externa/Adventitia (Fibroblasts & Collagen Layer)
  - GelMA/Collagen I (UV Light & CaCl<sub>2</sub>)
  - NIH 3T3 Fibroblast Cells
- Vessel Lumen
  - Pluronic<sup>®</sup> F127



### Tunica Intima (Endothelial Layer)

GelMA/Hyaluronic Acid (UV Light & CaCl<sub>2</sub>):

- GelMA
  - Biocompatible Biomaterial
  - High Mechanical Strength
  - Can Blend with Other Gels to Increase Cell Viability
- Hyaluronic Acid
  - $\circ$  Favorable Rheological Performance (Si et *al*. )
    - Following modification with methacrylic anhydride and crosslinking with 3,3'-dithiobis(propionylhydrazide)

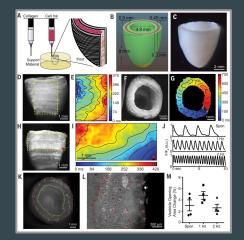
#### HUAEC/HUVEC Cells

- Co-Culture of HUVECs and HUAECs (Lau et *al*.)
  - Demonstrates optimal development of endothelium
- Mesenchymal Stem Cells (MSCs)
  - Substantial capacity in facilitating cardiac and endothelial repair

### Tunica Media & Externa/Adventitia

- Smooth Muscle Layer
  - GelMA/Alginate/PEGTA (UV Light & CaCl<sub>2</sub>)
    - Alginate is bioactive; tunable properties
    - PEGTA encapsulate cells
    - Vascular Smooth Muscle Cells
      - Characteristic of Layer
  - Mesenchymal Stem Cells
  - PEGTA
    - Cell Encapsulation
      - Prevents damage to cells in the process of printing

- Tunica Externa/Adventitia
  - GelMA/Collagen I (UV Light & CaCl<sub>2</sub>)
    - Collagen I has demonstrated use
    - NIH 3T3 Fibroblast Cells
      - Characteristic of Layer



## **Microfluidics**

### **Design Parameters for Microfluidic Chip**

#### Microfluidic Chip Material Microfluidic Chip Design

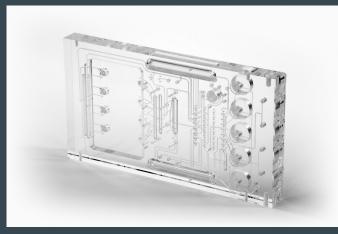
- PDMS (Polydimethylsiloxane)
  - Transparent
  - Forms Tight Seals with Plasma Treatment
- Easy to Mold
- Inexpensive
- Gas Permeable

Susceptibility of bubble formation and slight evaporation avoided through bubble traps

• Combination of Continuous and Droplet-Based Flow

- Incorporate 3D Hydrogels
- Bubble Trapper

Allows for the visualization of flow through fluorescence



Component(s)

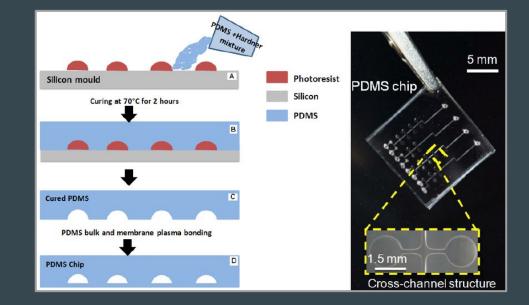
### **Microfluidics: Size and Fabrication**

#### • Size

- At least one dimension <500 μm
- Volume scale: 0.01 nL 1 L

#### • Fabricate with PDMS

- $\circ$   $\,$  Mold Master with CAD software  $\,$
- Contact Photolithography
- Glass Posts Placed Upright
- PDMS is Cast Against Master
- Plasma Oxidation for Sealing



### **Microfluidics: Module-Based Design**

#### Four Modules:

#### 1. Solutions

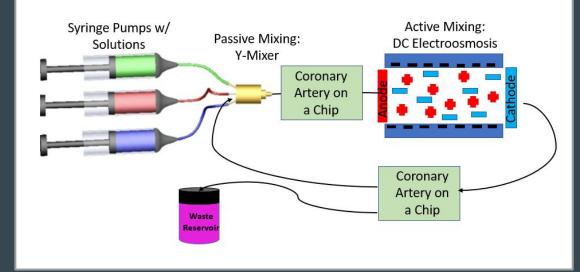
- a. Syringe Pumps with Blood and Nutrients
- b. Drop-Based

#### 2. Passive Mixing

- a. Diffusion
- b. Combo Y-Mixers
- c. Laminar Flow
- d. Connected to Syringes Supplying Solutions

#### 3. Active Mixing

- a. DC Electroosmosis Pump
- b. Pulsatile Flow
  - Goes to Arteries
- 4. Waste Reservoir



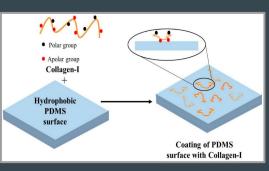
### **Microfluidics: Integration**

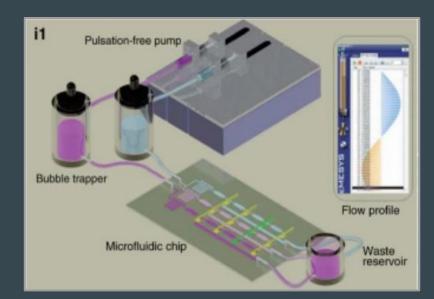
#### • Incorporation of 3D Hydrogels

- Collagen mimics cardiac or vascular tissue
- Correlation between decreased lymphatic
   barrier function and increased collagen
   density (Lugo-Cintron et *al*.)

#### • Bubble Trapper

- In-Line
  - Bubbles float to the top when they enter the network

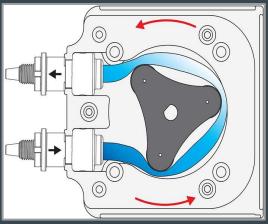




### **Microfluidics: Pulsatile Flow**

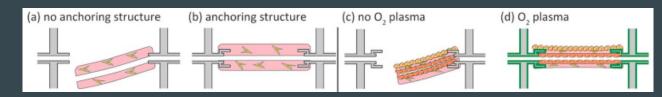
- Pulsatile Flow: Simulate cycles of contraction and relaxation in cardiovascular system
- Two Potential Methods
  - Varied Diameter Channels
    - 600 μm wide (healthy human artery) gradually narrows to 300 μm
    - Pulsatile shear stress applied: flow rate 1.41 µl/s at a pulse rate 70 bpm
    - Results in shear stress 15 dyn/cm<sup>2</sup> in 600 um channel (typical in healthy human arteries)
  - Peristaltic Pumps
    - Allows for continuous circulation of fluid and recirculations
    - Mechanism: pinches fluidic tube with roller on a rotor in a circular pump
    - Generate strong pulses
    - Non-fluidic contact operation

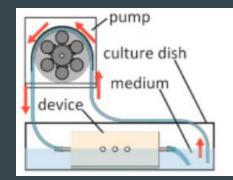


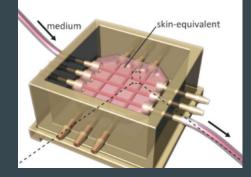


### **OOC: Integration of Microfluidic Chip & Bioprinted Coronary Arteries**

- Bioprinted coronary arteries within culture device (Mori et *al*.)
  - Both edges of the vascular channels fixed to connectors of culture device
  - Attached to a perfusion system composed of a peristaltic pumps and silicone tubes connected to passive Y-mixer
  - Culture device attached to anchoring structures with oxygen plasma treatment

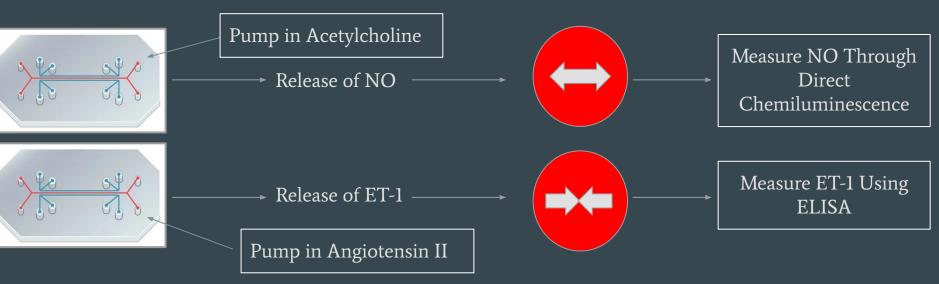






#### Testing and Validation: Endothelial & Smooth Muscle Cell Function

- Vasodilation and vasoconstriction are important functions of these cells
- Caused by the release of several factors (nitric oxide (NO), endothelin (ET-1))
- Commonly used test is to inject acetylcholine and angiotensin into the arteries



#### Testing and Validation: Flow, Permeability, Fibroblast Function

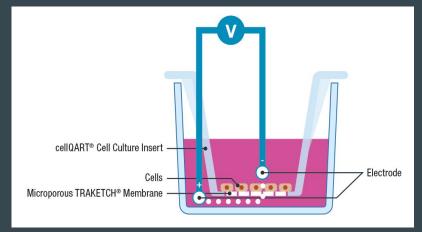
<u>Fibroblast</u>: Angiotensin II (Ang II) has shown to causes fibroblasts to produce inflammatory cytokines

• Pump in Ang II and measure cytokine production using multiplex ELISA

Pulsatile Flow: Use embedded sensors

- Average Flow: 1.41 μL/s
- Pressure > 1.5 Pa
- Average Shear Stress: 15 dyn/cm<sup>2</sup>

<u>Permeability</u>: Use transendothelial electrical resistance (TEER) to make sure there is no leakage



### **Biocompatibility**



- PDMS: Biocompatible material made from the repetition of silicon and oxygen bonds and methyl groups with low cost.
- GelMA has great biocompatibility and mechanical properties. It is developed by gelatin, the product of hydrolysis and collagen denaturation at high temperatures
- Hyaluronic Acid, Pluronic<sup>®</sup>F127, polyethylene glycol polyacrylamide copolymers (PEGTA) as hydrogels are all biocompatible material

#### **Application: Drug Testing and Efficacy**

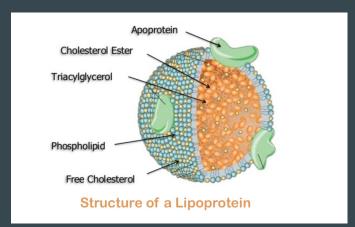
This device could be a way of testing drug efficacy in treating atherosclerosis without having to use an *in vivo* model - Alendronate used for osteoporosis

Create atherosclerosis condition

- Introduce **LDL** through shear stress when seeding endothelial cells
- Increase HR

Implement drug treatment and observe

• Pump in **alendronate** in desired dosage and see if the amount of plaque reduces or if the rate of plaque buildup decreases

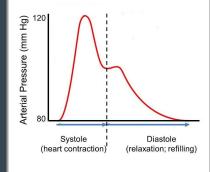


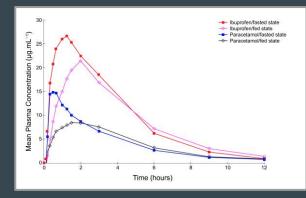
#### Limitations of Proposed OOC Technology for Coronary Arteries

- 1. Simplification of Pumping Mechanism
  - a. Pressure gradient along the coronary arteries is variable
  - b. Varied pressure gradient (and velocity) along the arteries can contribute to vascular insult

#### 2. Variable Composition of Fluid Over Time

- a. Multi-organ interactions result in varied composition of molecules and cells within blood as a function of time
- Certain molecules can increase or decrease likelihood of vascular insult
  - i. Over or under estimation of plaque build-up may result
- 3. 3D Bioprinting is a Relatively Novel Technology
  - a. Poses accessibility/scalability risk over current PDMS REM methods that are currently more utilized





# **Questions?**

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