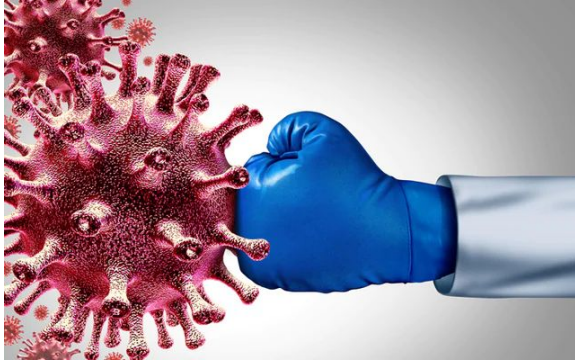


Hayden, Stef, Kira, and Luke

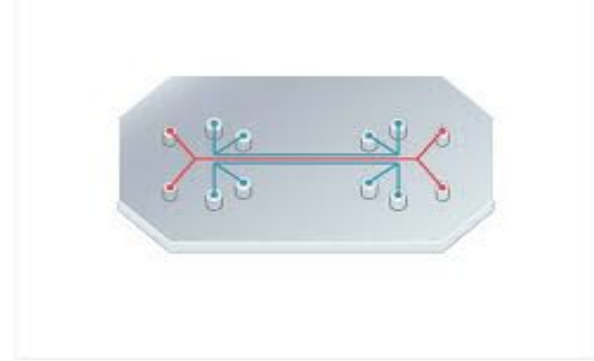


Thymus on a Chip

Background



Immune System



Organ on a Chip

Immune System

Immune Organs

Primary

Generate Immune Cells

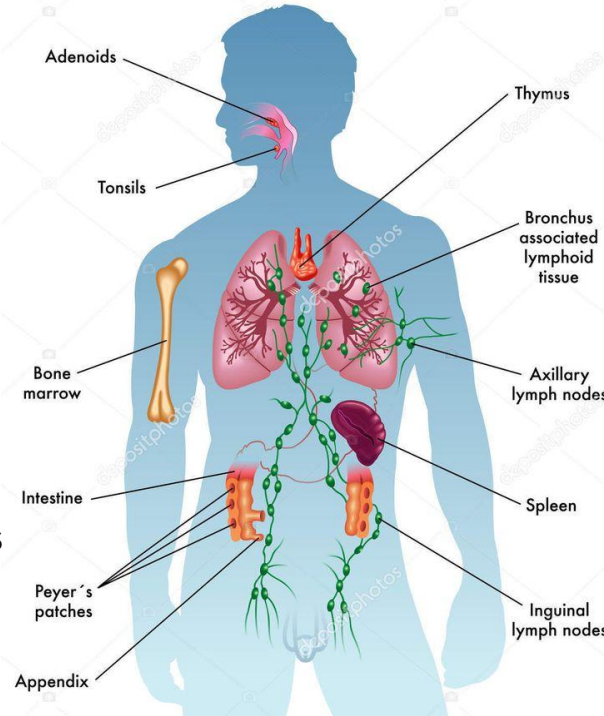
-Bone Marrow
-Thymus

Secondary

Cell programming

-Tonsils
-Lymph Nodes
-Spleen

Organs of the Immune System



Immune Cells

Myeloid Precursor

Eosinophils
Basophils
Neutrophils

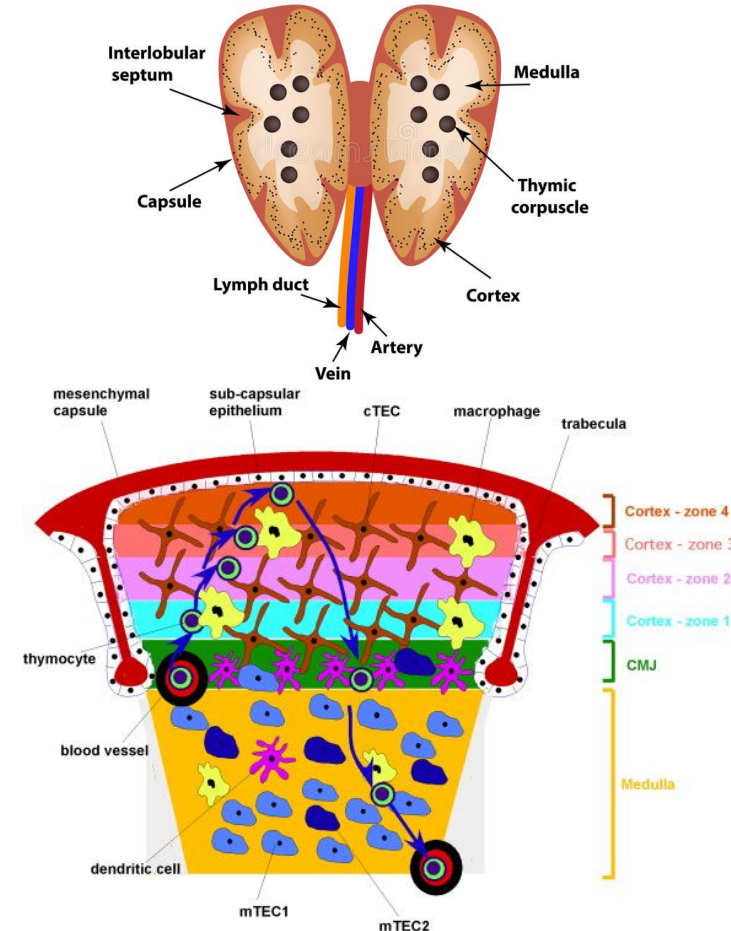
Lymphoid Precursor

T and B Lymphocytes

Thymus

Generates a diverse army of T cells essential to the efficient function of the adaptive immune system.

- Components:
 - Cortex and medulla in which two epithelial cell types reside:
 - cTECs and mTECs
 - Stromal cells: ECs, fibroblasts, and APCs.

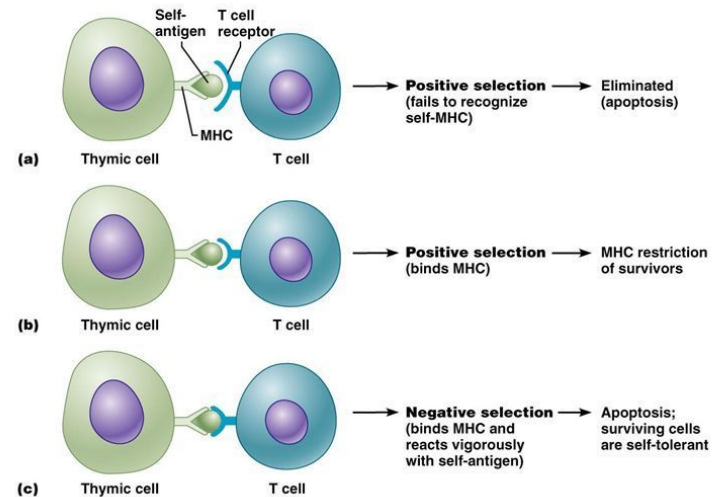
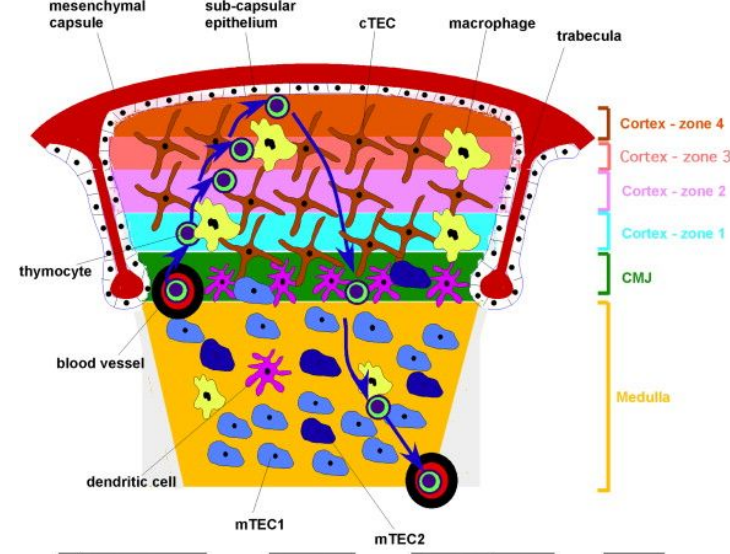


Thymus

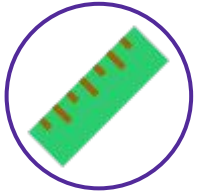
Main function: T cell selection and maturation

- Positive selection:
 - Ensure stable TCR interactions with cTECs.
 - Occurs within cortex
- Negative selection:
 - Ensure T-cells with TCRs for self-antigens are eliminated.
 - Occurs within medulla

These events require interactions with the TECs in the 3D stromal environment.



Current Challenges in Studying the Thymus



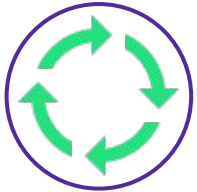
Scale

Limits Architectural Complexity

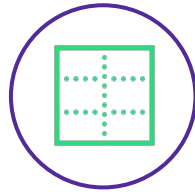


Accurate Representation of All Functions

Dependent on Architecture



Limited TEC Regenerative Capacity

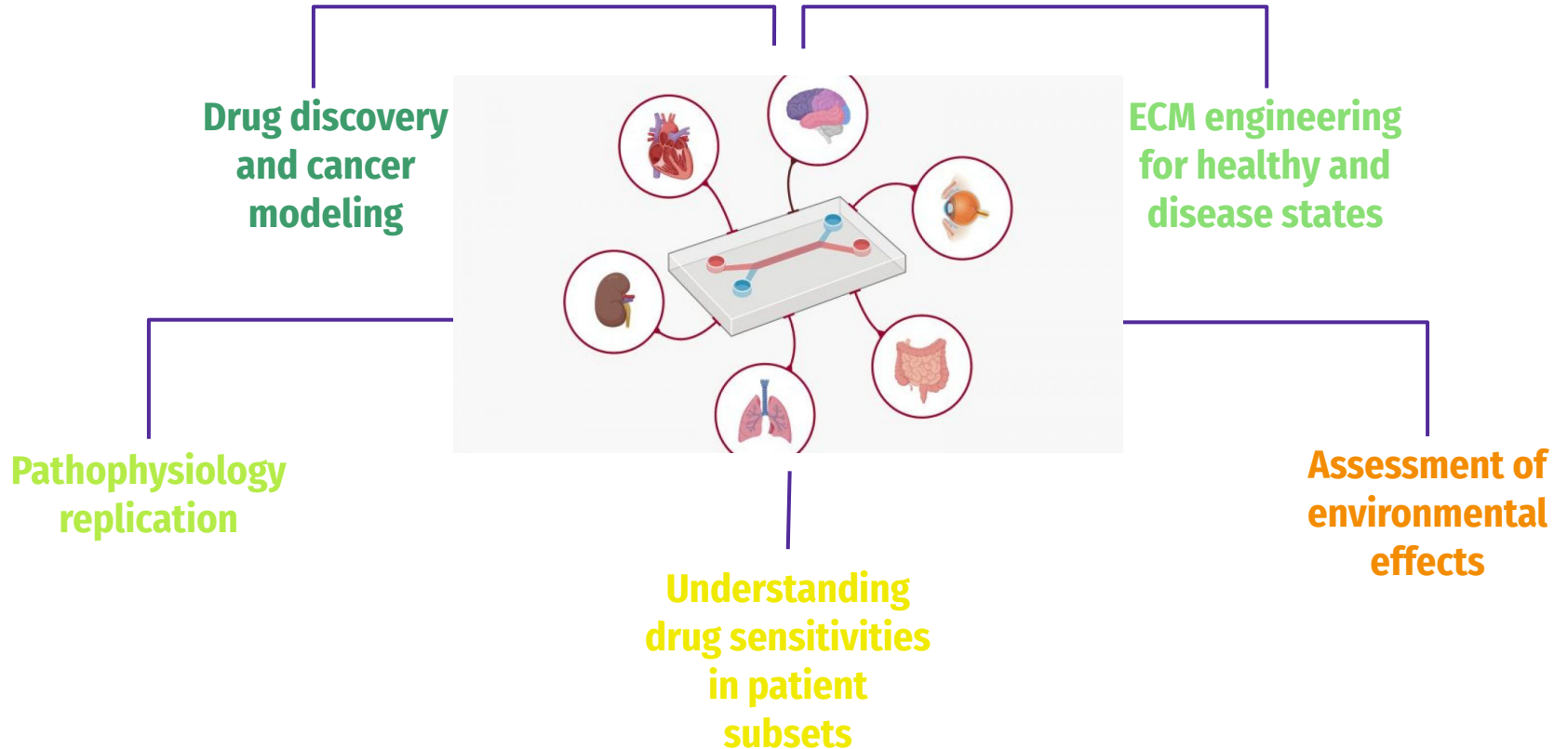


Current TEC Seeding Methods

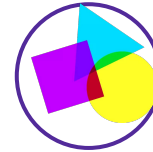
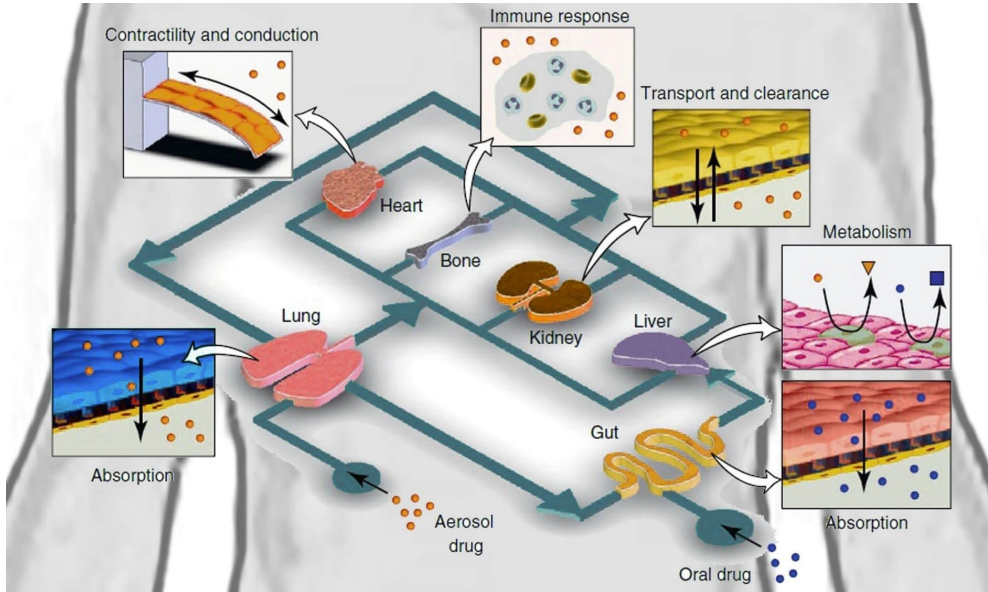
Poor Cell Viability



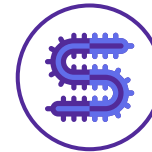
Organ-on-a-chip (OOAC)



OOAC Components

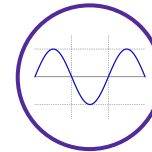


Geometric confinement and patterning



Control of flow

Inlets and outlets



Environmental control

Mechanical/electrical stimulation

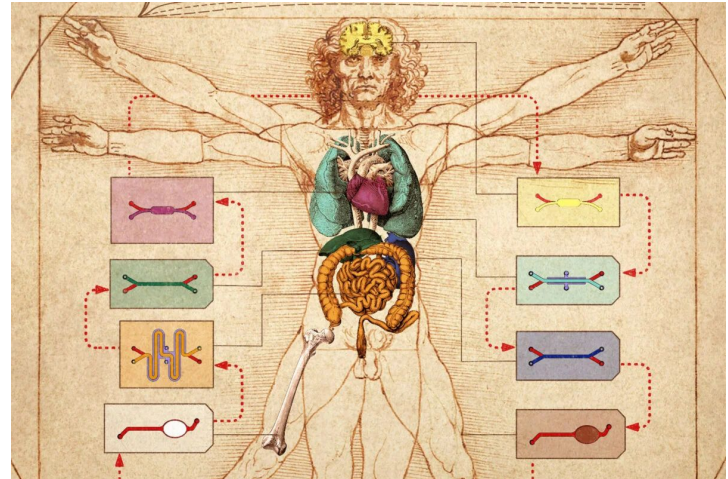


Sensors

Need for Immune Response in OOAC

“So far, the development of OoC systems to emulate the immune system has lagged behind”

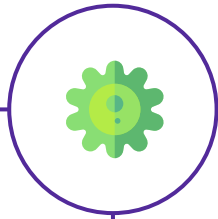
- Necessary for complete body-on-a-chip
- Critical to fully characterize/test new drugs
- Complete pathophysiologies cannot be replicated without immune response



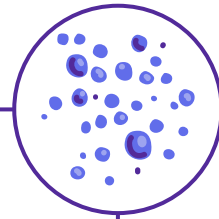
Theory and Principles



Cell Culture



ECM
Reconstruction



Microfluidics



Cell Culture

Options for Labs

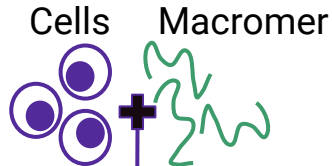
1. Purchase a cell line (VECs)
2. Harvest cells from animal (TECs)

Reaggregate Thymic Organ Cultures

- Harvest thymic lobes → cut into small pieces → filter thymic remnants → select and separate desired cells (TECs)
- Continue to culture and split cells as needed



ECM Reconstruction



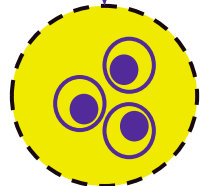
Cell laden hydrogel network



Secretion of matrix,
Hydrogel degradation



Functional tissue



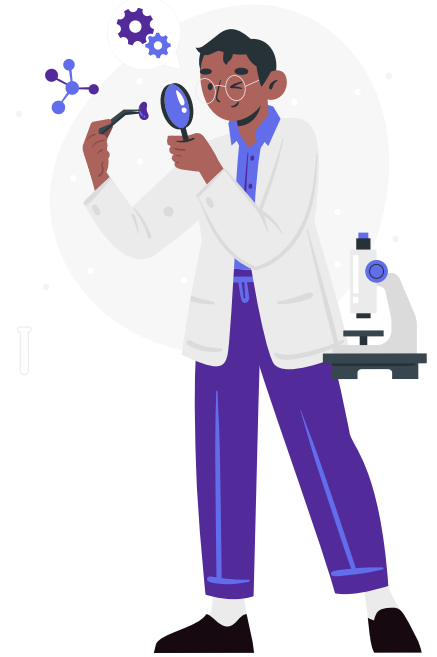
ECM Function:

- Cell adhesion
- Cell growth
- Cell signaling

ECM Reconstruction

- Hydrogel

Microfluidics



→ Features less than 1 mm & internal volumes less than 100 μL

Pros:

1. Less volume and analyte required
2. Gravity and inertia become less relevant at this scale

$$\text{Re} = \frac{\rho V D}{\mu}$$

Small diameters → Small Reynolds numbers
→ **Laminar flow** ✓

$$\bar{x}^2 = 2Dt$$

In laminar flow, mixing is predominately by **diffusion**

Requirements of Modeling the Thymus

1.

Utilization of functional cell types of the Thymus

2.

Suitable biomaterials for cell adhesion/cell interactions

3.

Cell viable environment

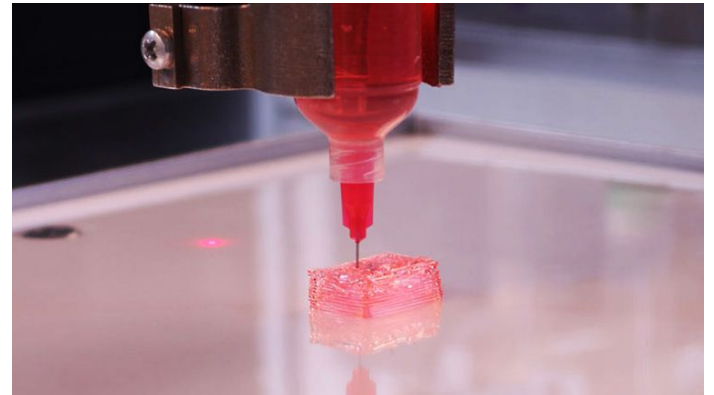
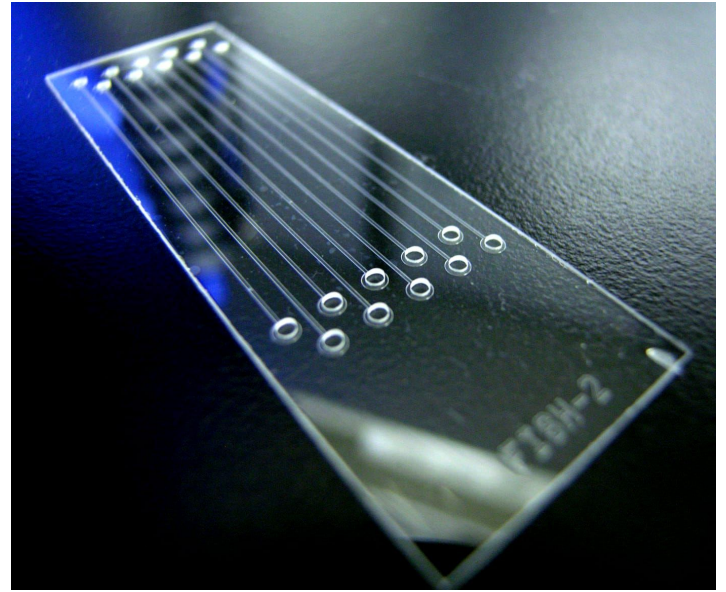
4.

Monitoring of T-cell selection

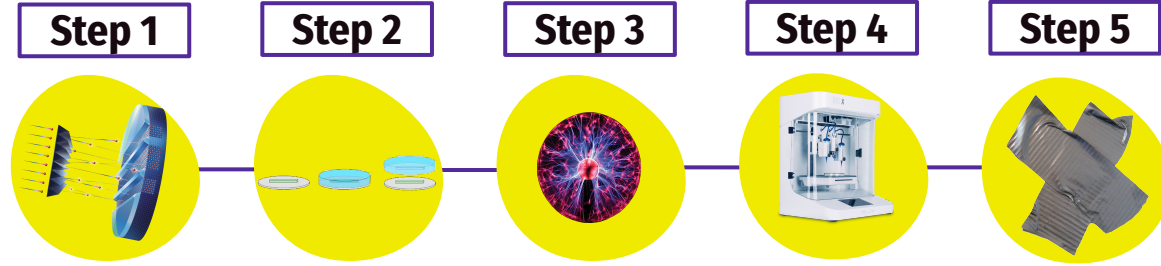


Device Fabrication

Microfabrication
+
3D Bioprinting



Microfluidic Fabrication



Develop Si Wafers

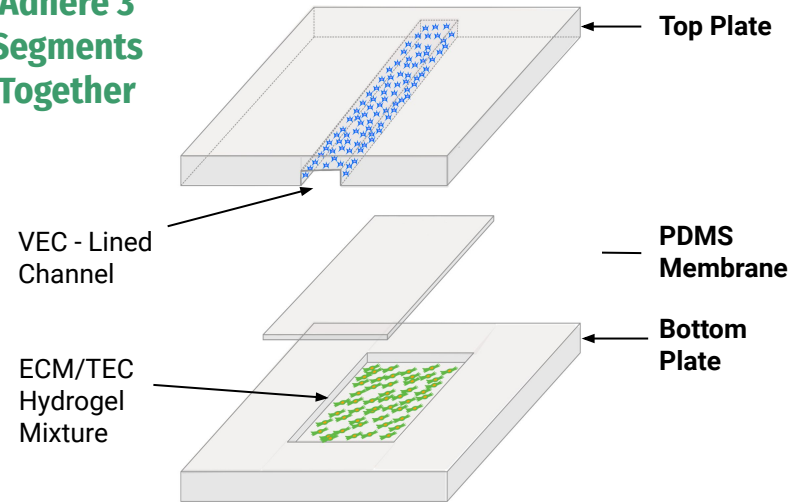
Cast PDMS

O₂ Plasma Treatment

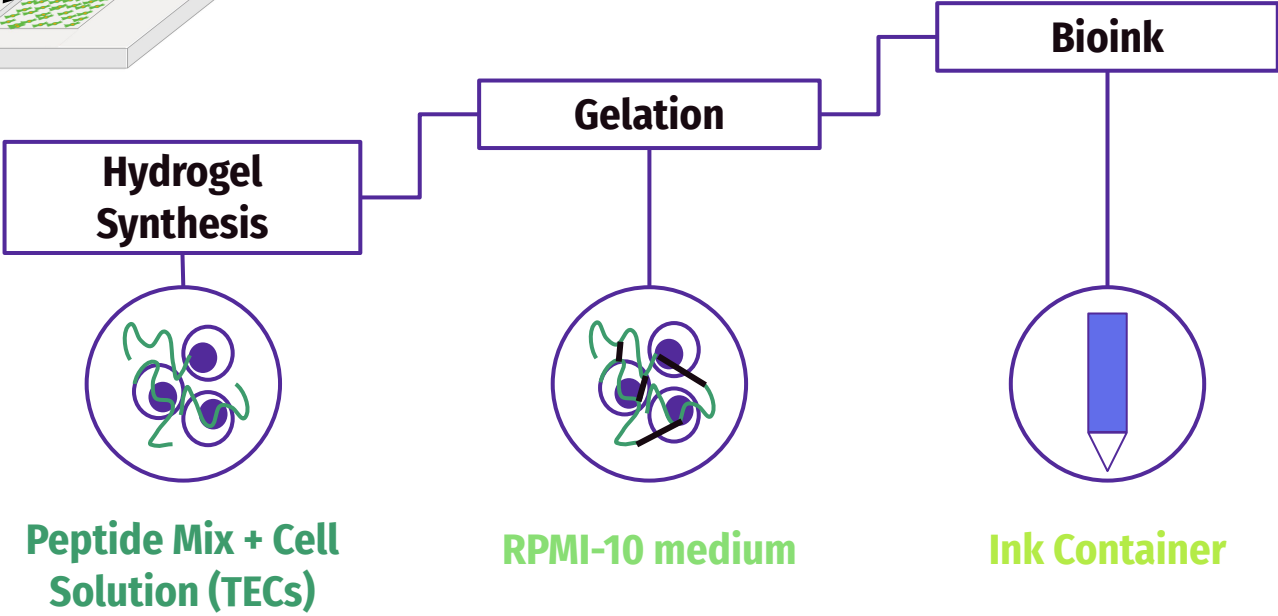
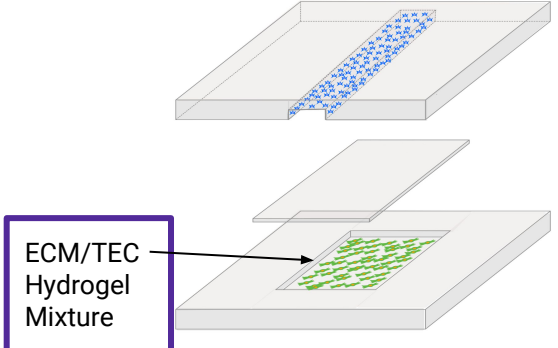
Bioprint Functional Unit

Adhere 3 Segments Together

- 1) Top Microfluidic Channel
- 2) Thin Porous Membrane
- 3) Bottom Functional Unit Pocket



Bioink Construction



Endothelialization of Channels w/ VECs

Why is this component needed?

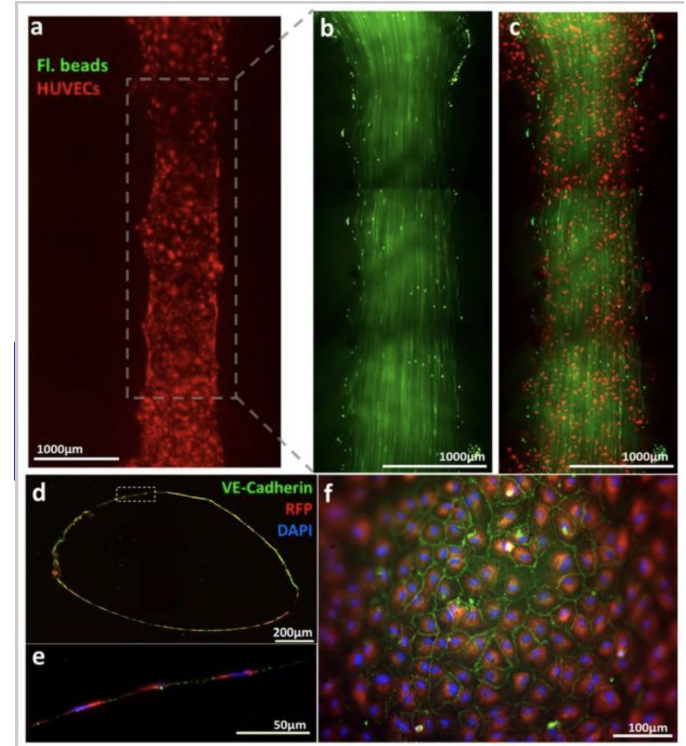
- Current thymus organoids do not include vasculature

Deposition of Adhesive ECM Components

- Fibronectin coating

Cell Suspension

- Connect tubes to channel inlet and outlet
- Flow VEC solution through
- Place on rocker for 5 minutes
- Flip upside down



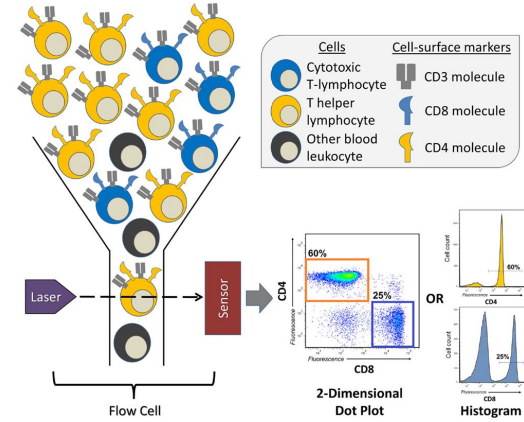
Testing & Validation



Biosensing

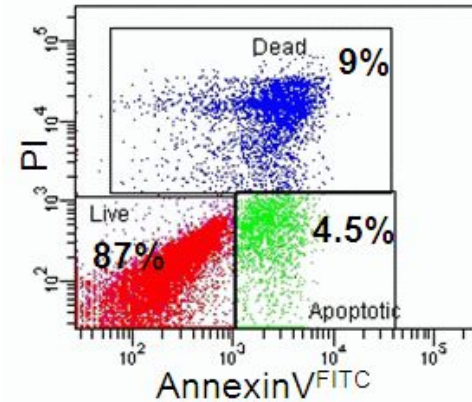
Stain appropriate cell surface markers indicative of T cell maturation

- CD3+: All T Cells
 - CD8+: Cytotoxic
 - CD4+: Helper
 - CD25+: Regulatory

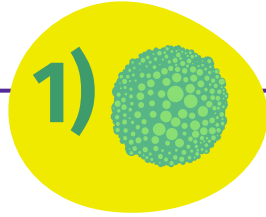


Cell Viability: Annexin V (Stains PLs) and Propidium Iodide (Membrane Integrity)

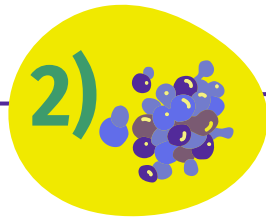
- Live Cells => PL within cell
- Apoptotic => PL exposed
- Other Death => PI stained



Flow Cytometry



1) (+) Selection and Successful Maturation

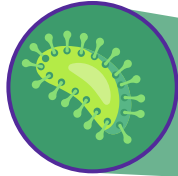


2) (-) Selection

Validation

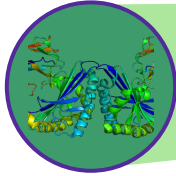
Cell Viability

Are the cells alive post-implantation?



Testing loss of TECs

Are epithelial cells being lost when solution is flowed?



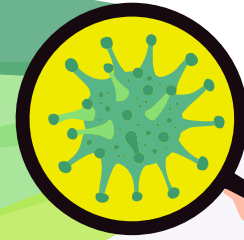
Expected Function

Are the T-cells being "killed" as expected?



User Interviews

Will the device be useful?



Cell Viability

Hydrogel Cell Culture

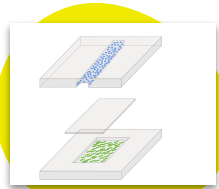
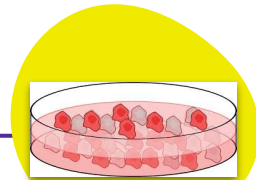
Fluoroprobe CMFDA

Bind fluoroprobe CMFDA to cells
Place cells into hydrogel
Use computer software to measure the amount of viable cells with an image

Liquid Cell Culture

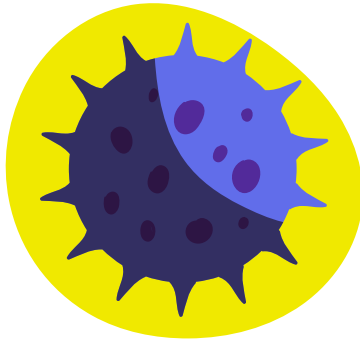
Trypan blue staining

Stain cells with trypan blue
Implant
Collect sample after 1 hour, 1 day, 1 week, and 2 weeks
Use a hemocytometer and measure amount of viable cells



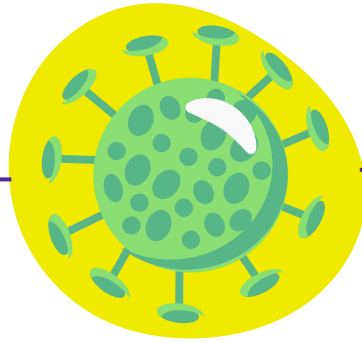
TOAC (Thymus on a chip)

Quantifying Loss of TECs



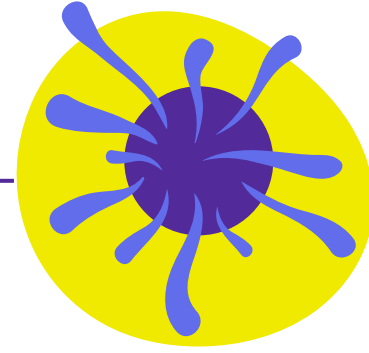
Finalize Prototype

Finalize hydrogel and complete prototype



Flow T cell solution

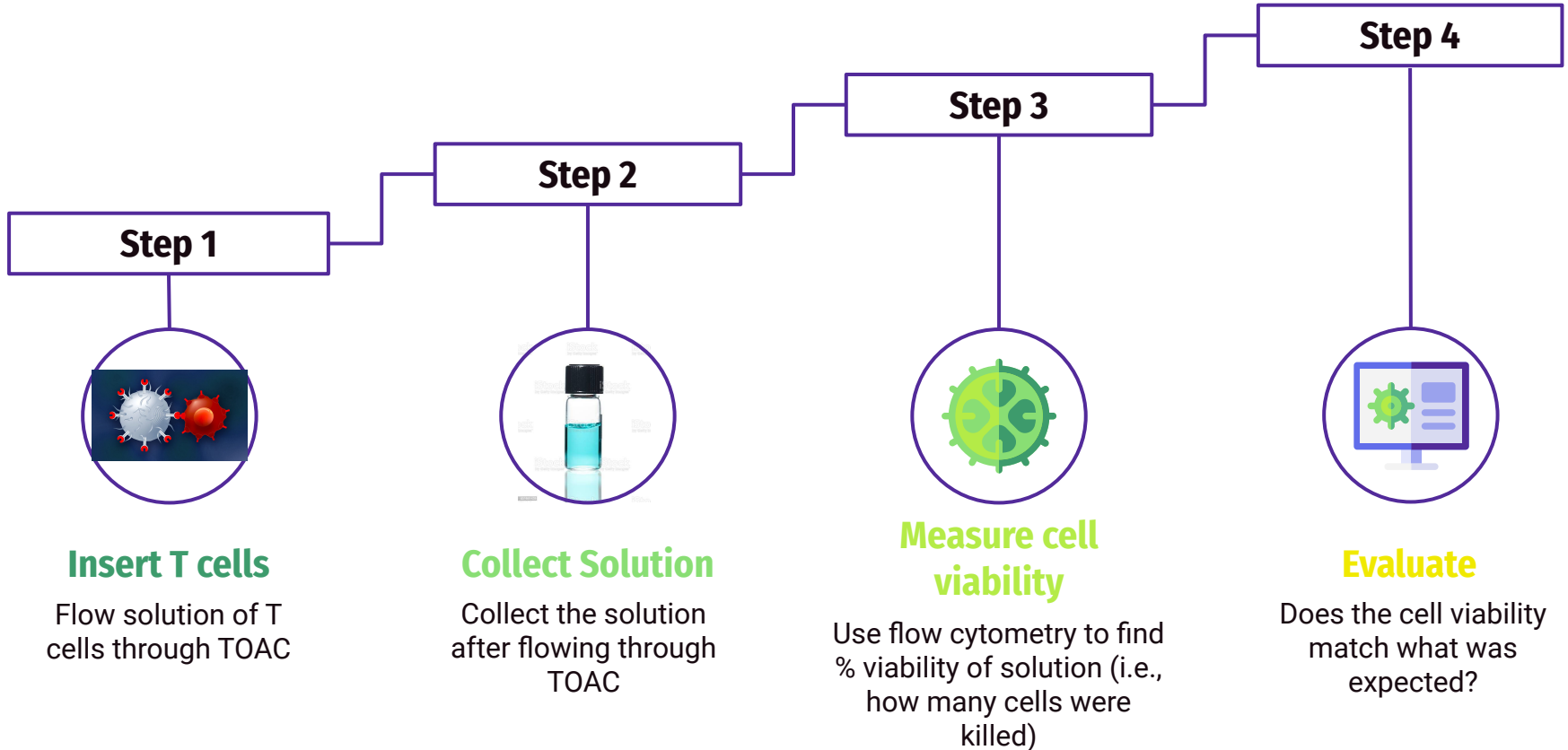
Flow solution of T cells through device



Flow cytometry

Add stain to solution to use flow cytometry and ensure no stained CD1a leaves the device

Expected Function



User Interviews



Rate the usefulness

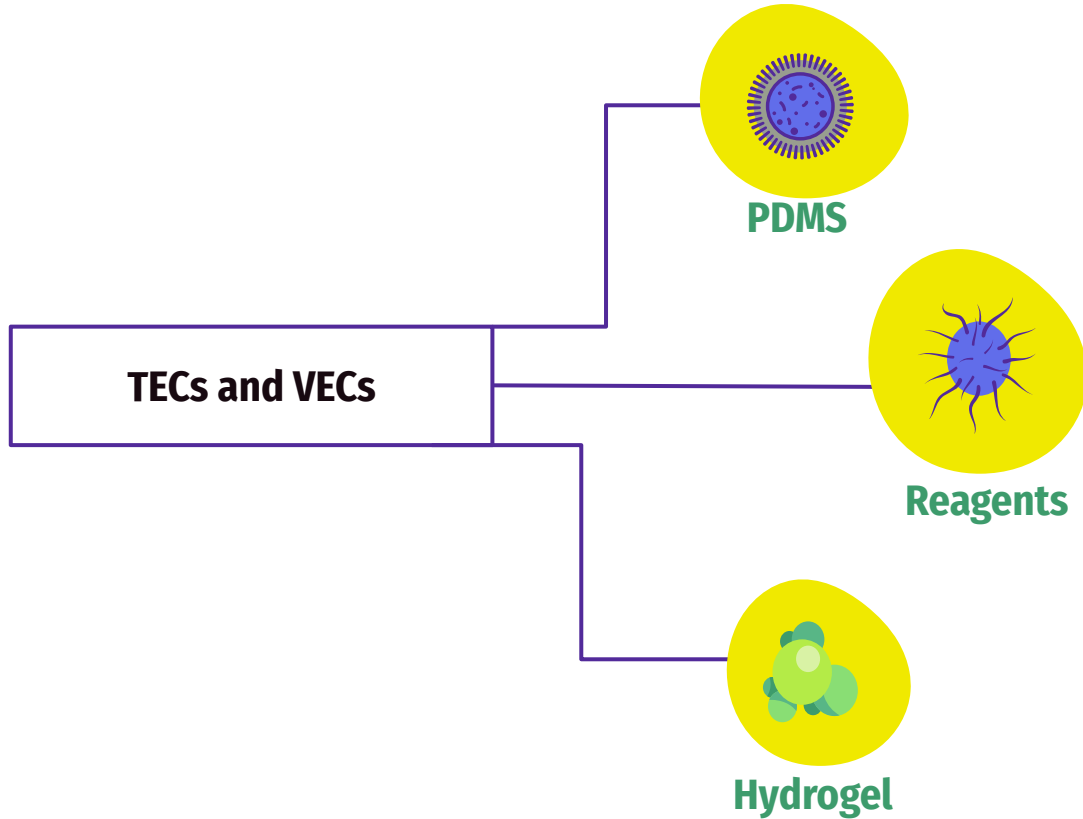


Rate the usability

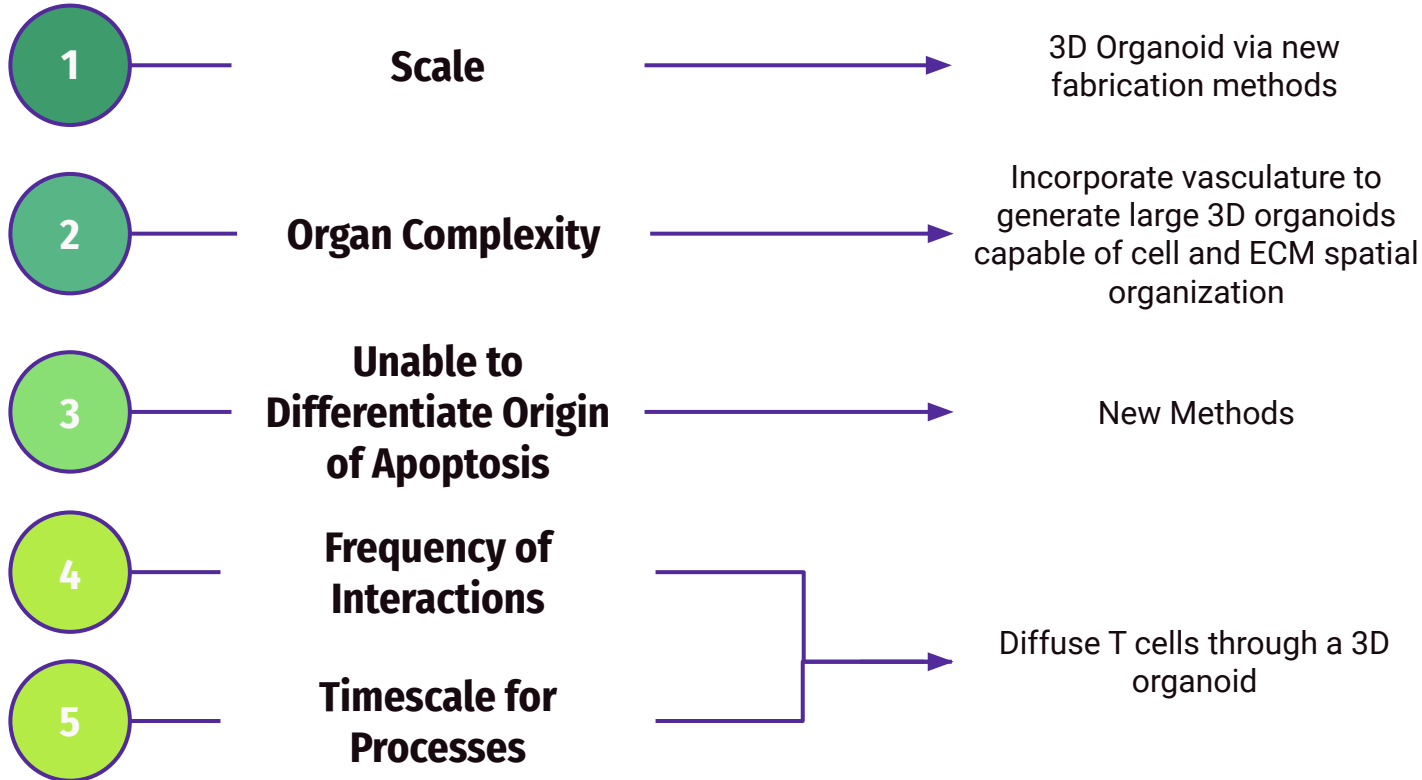


What is missing that you would like to see?

Biocompatibility



Limitations → Future Directions



References

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7. Deng Z., Liu H., Rui J., Liu X. (2016) Reconstituted Thymus Organ Culture. In: Bosselut R., S. Vacchio M. (eds) T-Cell
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Yong Fan, Asako Tajima, Saik Kia Goh, et al. Bioengineering Thymus Organoids to Restore Thymic Function and Induce Donor-Specific Immune Tolerance to Allografts, *Molecular Therapy*, Volume 23, Issue 7, 2015, Pages 1262-1277.
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- 10 Vijayasekaran, S. "Cell Viability and Inflammatory Response in Hydrogel Sponges Implanted in the Rabbit Cornea." *Biomaterials*, vol. 19, no. 24, 1998, pp. 2255–2267., [https://doi.org/10.1016/s0142-9612\(98\)00128-8](https://doi.org/10.1016/s0142-9612(98)00128-8).
- 11 Riss T, Niles A, Moravec R, et al. Cytotoxicity Assays: In Vitro Methods to Measure Dead Cells. 2019 May 1. In: Markossian S, Grossman A, Brimacombe K, et al., editors. *Assay Guidance Manual* [Internet]. Bethesda (MD): Eli Lilly & Company and the National Center for Advancing Translational Sciences; 2004-. Available from: <https://www.ncbi.nlm.nih.gov/sites/books/NBK540958/>
- 12 De Jong, Annemieke, et al. "CD1A-Autoreactive T Cells Are a Normal Component of the Human A β T Cell Repertoire." *Nature Immunology*, vol. 11, no. 12, 2010, pp. 1102–1109., <https://doi.org/10.1038/ni.1956>.

Questions?