

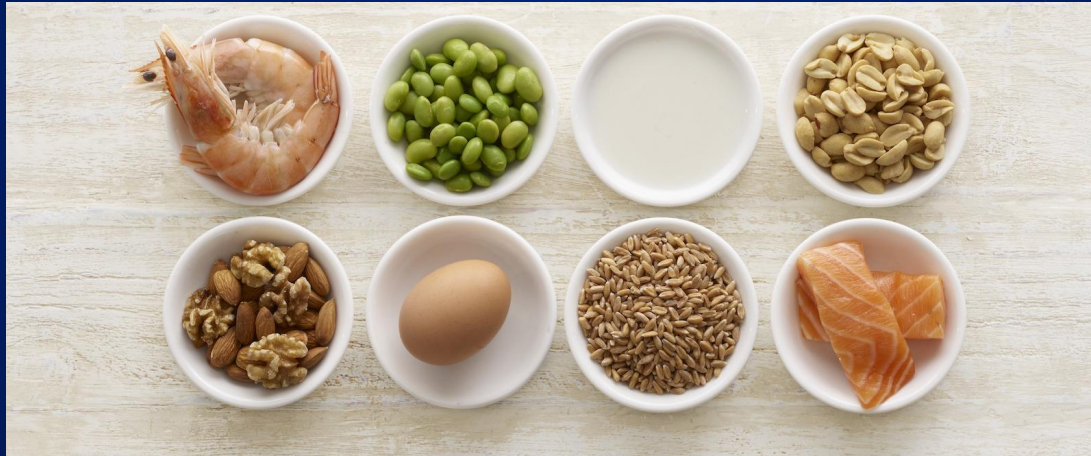


Gut-On-A-Chip to Test for Food Allergies and Intolerances

Annika, Melissa, Nadeen, Taylor

Background

- 32 million people in the US suffer from food allergies
- Food allergies cause immune system dysfunction and painful digestion
- Many people with food allergies try elimination diets
- Lots of failure due to timeliness and difficulty



Current Gaps

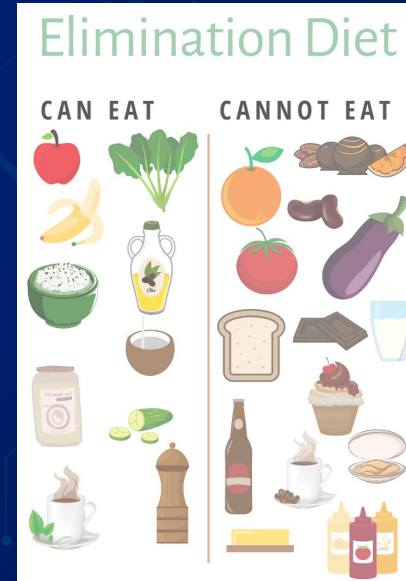
Food Sensitivity Tests

1. Blood work required
2. Does not give good insight into overall digestive dysfunction

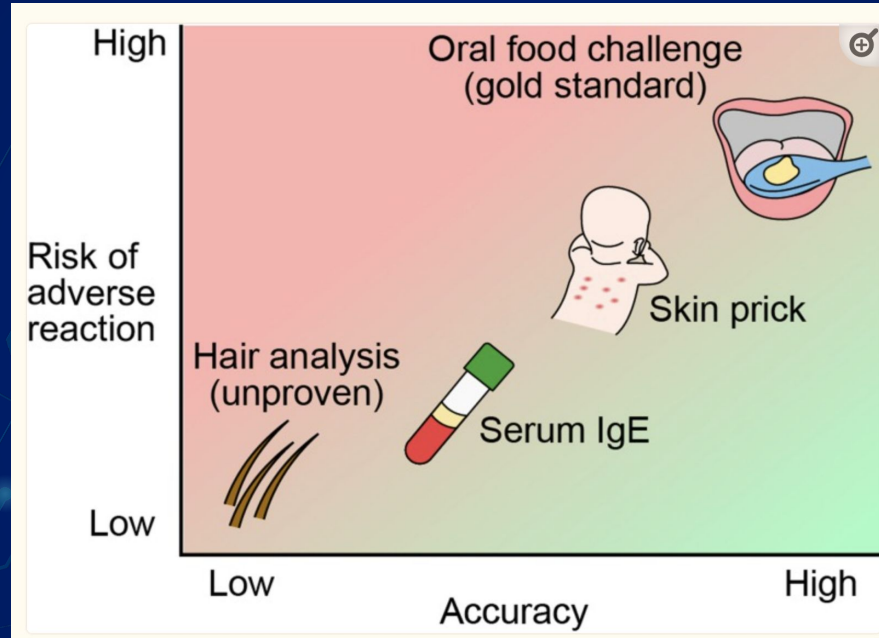


Elimination Diets

1. Time-consuming
2. Difficult to follow through with



Current common allergy testing methods present a trade off between accuracy and risk of adverse reactions.



Castaño, Nicolas, et al. "Microfluidic Methods for Precision Diagnostics in Food Allergy." *Biomicrofluidics*, vol. 14, no. 2, Apr. 2020, p. 021503. PubMed Central, <https://doi.org/10.1063/1.5144135>.

Key Features of the Gut

Transport

Absorb and transport nutrients from digestion to vasculature

Specialized Cells

Microbiomes to make vitamins and break down complex foods

Immunity

Trains the immune system and rids system of pathogens

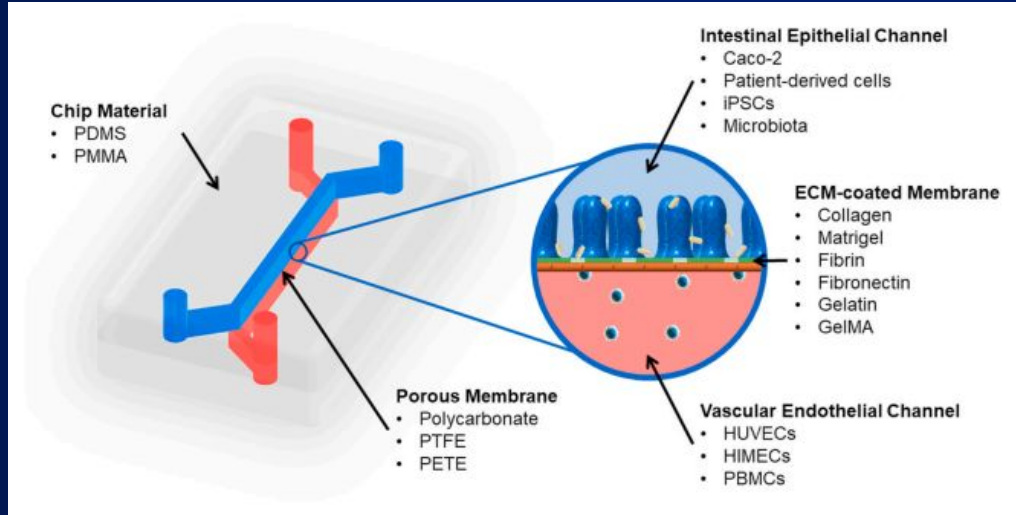
Structural

Intestinal villi and microvilli increase surface area for nutrient absorption

Mechanical

Peristaltic contractions move food through the GI tract

Existing Gut-On-A-Chip Models



- Channels and chambers
 - Sensors, electrodes, valves
- Gas permeable
- Vascular endothelial cells
- PDMS

Ashammakhi, N., Nasiri, R., Barros, N. R., Tebon, P., Thakor, J., Goudie, M., Shamloo, A., Martin, M. G., & Khademhosseini, A. (2020). Gut-on-a-chip: Current progress and future opportunities. *Biomaterials*, 255. <https://doi.org/10.1016/j.biomaterials.2020.120196>

Limitations with Current Technology

- Balance of complexity and relevance
 - Cost
 - Ease-of-use
- Incorporation of physiologically relevant cells
 - Currently use animal-derived or immortalized cell lines
 - Unable to accurately mimic *in vivo* biology
- Incorporation of biosensors and material that responds to stimuli
- Limited life span of cells on chip

Requirements for Modeling the Gut

**Cellular
composition**

**3D
Architecture**

**Dynamic flow
conditions**

Motility

Proposed Solution

A gut-on-a-chip system that can diagnose food allergies and sensitivities as well as show the physiological response within the gut with less pain and high accuracy

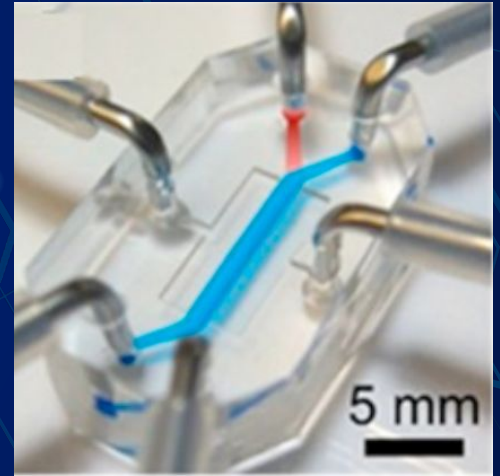
- **What the model will achieve:**
 - A vasculature of blood vessels that will help us detect the immune cells response
 - Real time measurements of live cells and subcellular processes
 - Real mimicking of intracellular gut environment
 -
- **Benefits**

Patient Specific

Time Effective

Less Painful

Predicts specific allergic reactions



Design Parameters

**Geometric
Confinement
& Patterning**

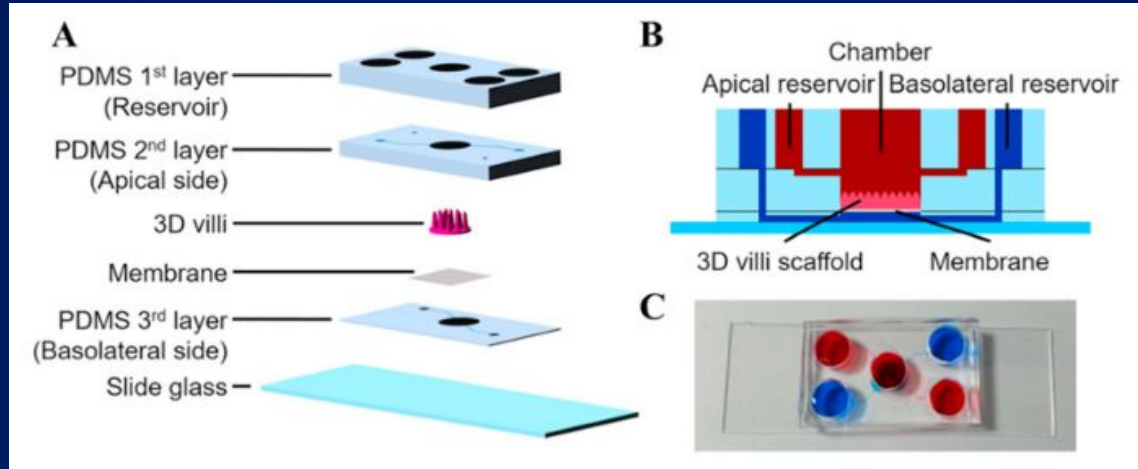
**Incorporating
cell samples
into the chip**

**Mimicking
real cell
environment
& ECM**

**Delivering
allergen into
the chip**

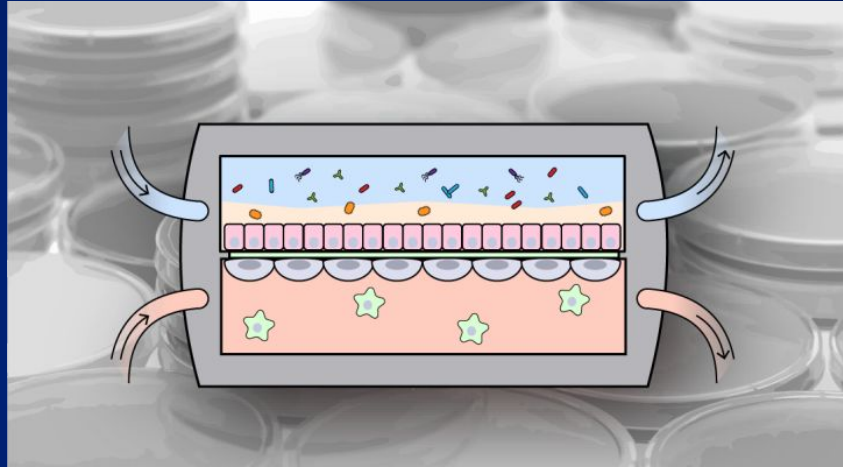
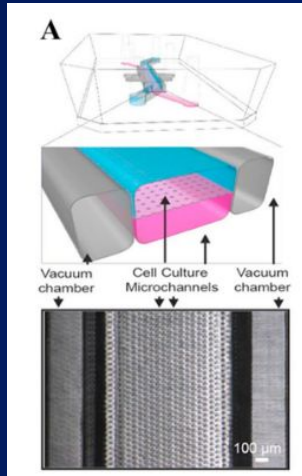
**Biosensing
and
physiological
readouts**

Geometric Patterning & Confinement



- Villi created using a dissolvable mold covered in collagen and seeded with cells (current studies use Caco-2 cells)
- Membrane composed of porous PDMS (0.45 μm pore size)

Geometric Patterning & Confinement

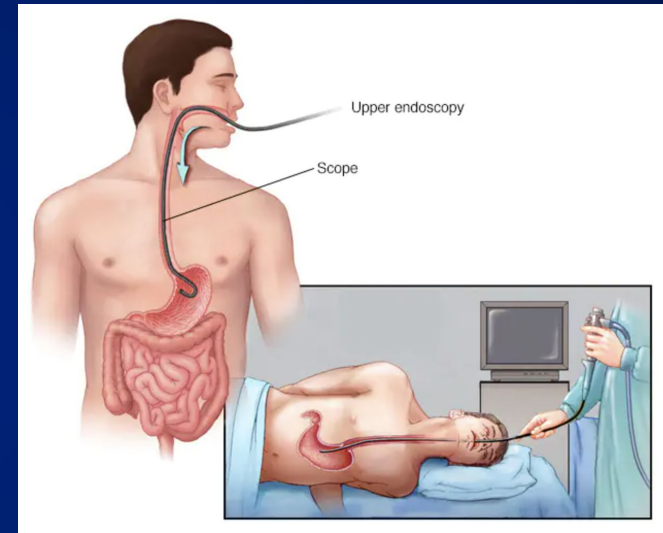
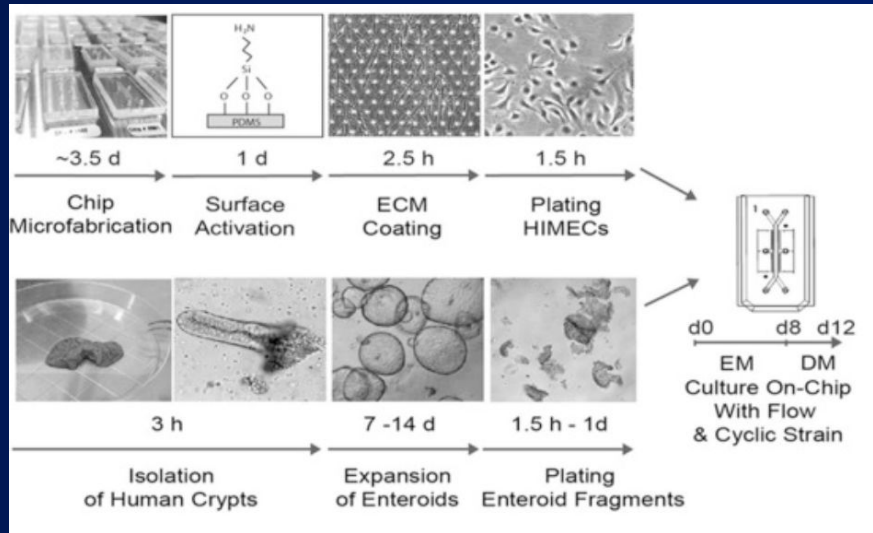


- Endothelial cells
- Vacuum chambers assist in peristaltic movement

Ashammakhi, N., Nasiri, R., Barros, N. R., Tebon, P., Thakor, J., Goudie, M., Shamloo, A., Martin, M. G., & Khademhosseini, A. (2020). Gut-on-a-chip: Current progress and future opportunities. *Biomaterials*, 255. <https://doi.org/10.1016/j.biomaterials.2020.120196>

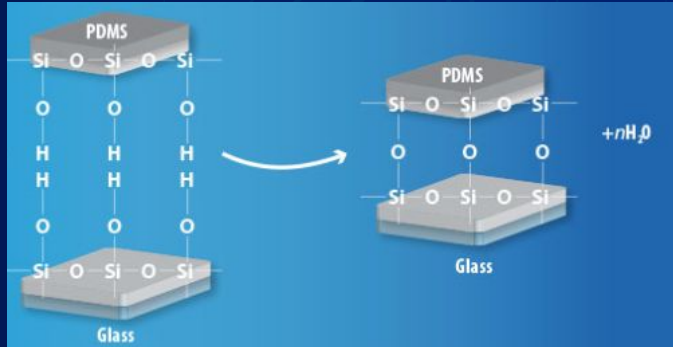
Elzinga, J. (2021, November 15). Towards a gut-microbiota-on-a-chip. WUR. Retrieved April 15, 2022, from <https://www.wur.nl/en/show/Towards-a-gut-microbiota-on-a-chip.htm>

Incorporating Cell Samples into the Chip

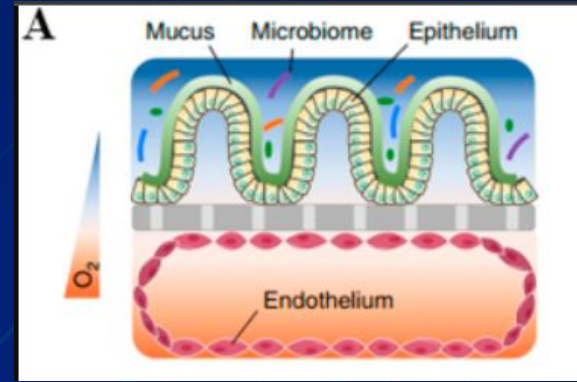


- Vascular Network
 - HUVECs or HIMECs
- Intestinal Network
 - primary human duodenal cells

Mimicking the Real Environment



- Treat the PDMS with O_2 plasma
- Increases hydrophobicity
- Decreases gas escape
- Lasts longer when kept and used in vacuum space


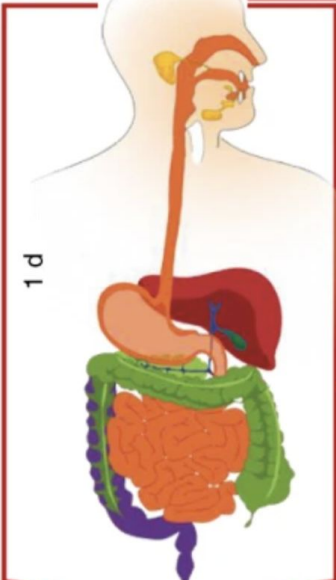


Other Features

- Lumen channel - patient's cells
- Vascular channel
- Microvilli
- Contractions

Delivering Allergens

- Need to digest the food that is being tested and then deliver this into the intestinal channel

 <p>5 d</p>	Preparation	<ul style="list-style-type: none"> • Perform enzyme activity and bile assays • Prepare SSF, SGF and SIF stock solutions • Perform pH-test adjustment experiment 	Step	<p>1</p> <p>2</p> <p>4</p>			
	 <p>1 d</p>	Oral phase	<ul style="list-style-type: none"> • Mix Food with SSF (1:1, (wt/wt)) • Include CaCl_2 (1.5 mM in SSF) • Add salivary amylase, if necessary (75 U/mL) • Incubate while mixing (2 min, 37 °C, pH 7) 	7–12	13	14	15, 16
		Gastric phase	<ul style="list-style-type: none"> • Mix oral bolus with SGF (1:1 (vol/vol)) • Include CaCl_2 (0.15 mM in SGF) • Add pepsin, gastric lipase (2,000, 60 U/mL) • Incubate while mixing (2 h, 37 °C, pH 3.0) 	17, 18	19	20, 21	22–24
		Intestinal phase	<ul style="list-style-type: none"> • Mix gastric chyme with SIF (1:1 (vol/vol)) • Include bile (10 mM bile salts) • Include CaCl_2 (0.6 mM in SIF) • Add pancreatin (trypsin activity 100 U/mL) • Incubate while mixing (2 h, 37 °C, pH 7.0) 	25, 26	27	28	29

BioSensing and Physiological Readouts

Biomarkers:

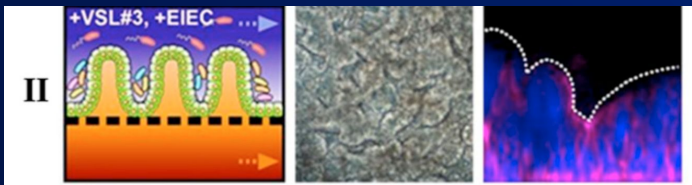
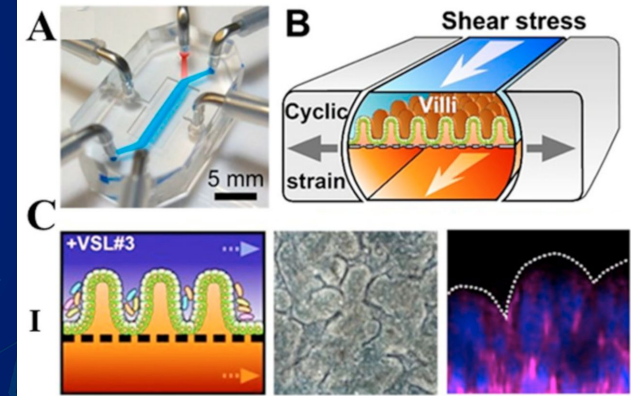
- There's not yet a single universal biomarker to diagnose food allergies.
- Ideal: detect the different biomarkers simultaneously to effectively track the disease

Cytokine Release and inflammation:

Immune cells produce proinflammatory cytokines in response to allergen presence

Cyclic mechanical strain and shear stress induced on the villi due to inflammation

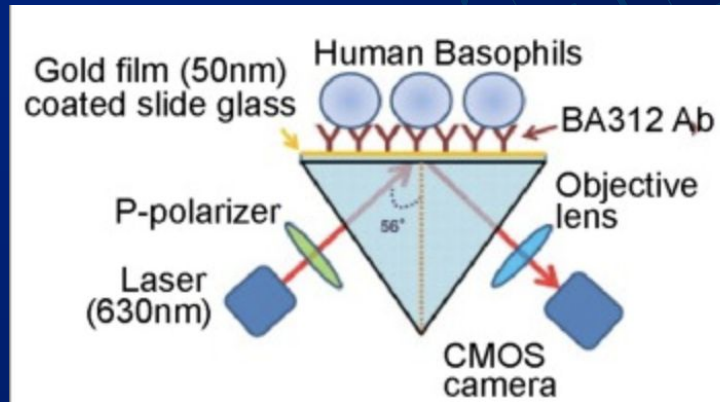
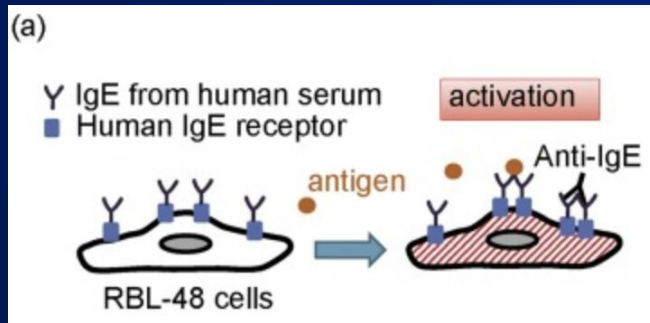
Visualize: fluorescent micrographs and phase contrast images



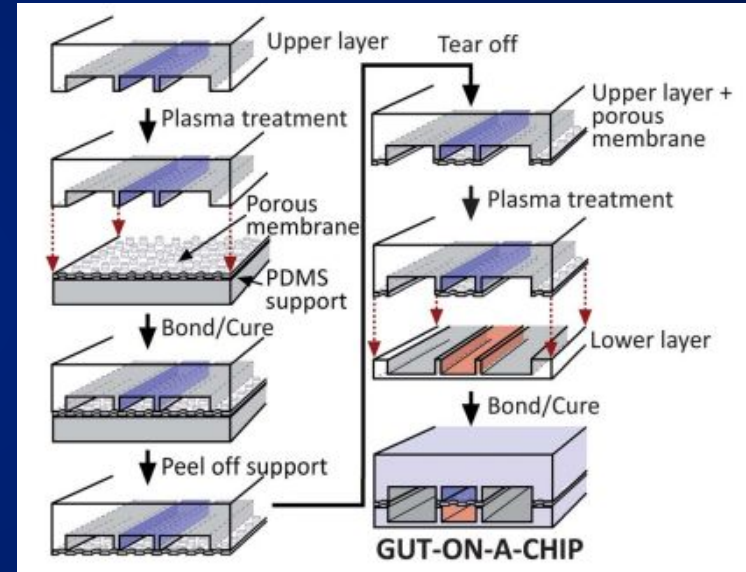
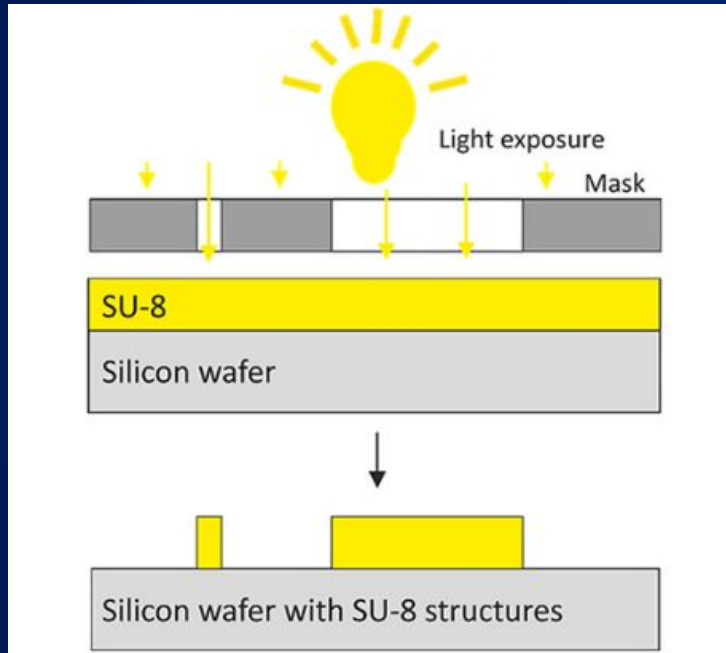
Ashammakhi, Nureddin, et al.

Surface Plasmon Resonance

- Examine sensitive changes to optical properties induced by intracellular changes like morphological changes and cell adhesion
- Cell degranulation
- Fix basophils with BA312 antibodies to the sensor surface and capture the change in intensity and in cell refractive index upon activation in real time.



Fabrication

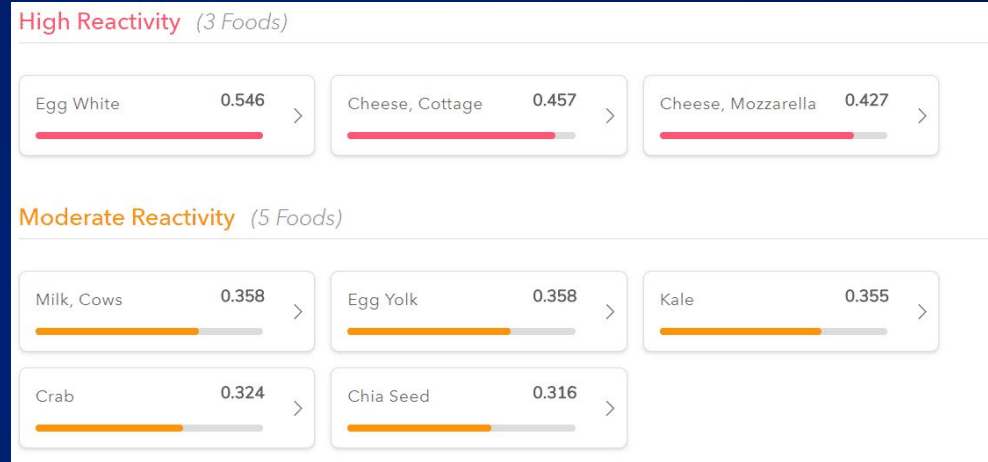


Elsbeth G.B.M. Bossink, Loes I. Segerink, Mathieu Odijk, Organ-on-Chip Technology for Aerobic Intestinal Host – Anaerobic Microbiota Research, *Organs-on-a-Chip*, Volume 4, 2022, 100013, ISSN 2666-1020, <https://doi.org/10.1016/j.ooc.2021.100013>.

Kim HJ, Huh D, Hamilton G, Ingber DE. Human gut-on-a-chip inhabited by microbial flora that experiences intestinal peristalsis-like motions and flow. *Lab Chip*. 2012 Jun 21;12(12):2165-74. doi: 10.1039/c2lc40074j.

Testing

EverlyWell Test Results



- Compare to food sensitivity tests or elimination diets
- Compare immune response to reactions from other allergens (same patient)
- Variety of tests to verify the results

Limitations

1. Sourcing cells from specific patients to test
2. Cost
3. Time to Manufacture
4. Does not account for interaction with other organ systems

Summary

- New gut-on-a-chip model to test for food allergies and intolerances
- Combine characteristics from existing chip models
- Specialize each chip to each patient



A light blue speech bubble is centered on a dark blue background. The background features a faint, glowing circuit board pattern with lines and nodes. The word "Questions" is written in a bold, italicized, black serif font inside the speech bubble.

Questions

References

1. Ashammakhi, N., Nasiri, R., Barros, N. R., Tebon, P., Thakor, J., Goudie, M., Shamloo, A., Martin, M. G., & Khademhosseini, A. (2020). Gut-on-a-chip: Current progress and future opportunities. *Biomaterials*, 255. <https://doi.org/10.1016/j.biomaterials.2020.120196>
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